

## SI22: ORF14

Table 1: General Information

<b>1</b>	<b>Protein Name</b>
	ORF14
<b>2</b>	<b>Region/Name/Further Specification</b>
<b>3</b>	<b>Sequence of fl protein</b>
	MLQSCYNFLKEQHCQKASTQKGAEAAVKPLLVP HHVVATVQEIQQA AVGELLLLLEWLAMA VMLLLLCCCLTD
<b>4</b>	<b>Protein boundaries of expressed construct</b>
	aa 1-73 (fl ORF14)
<b>5</b>	<b>Ratio for construct design</b>
	fl protein
<b>6</b>	<b>Sequence homology (to SCoV)</b>
	Identity: NA; similarity: NA
<b>7</b>	<b>Published structures (SCoV2 or homologue variants)</b>
	-
<b>8</b>	<b>(Published) assignment (SCoV2 or homologue variants)</b>
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Table 2: Cell-free Protein Synthesis

<b>1</b>	<b>Expression vector</b>
	pEU-E01-MCS (Cell-Free Sciences)
<b>2</b>	<b>Purification-/Solubility-Tag</b>
	N-terminal Strep tag II (WSHPQFEK)
<b>3</b>	<b>Cleavage Site</b>
	-
<b>4</b>	<b>Molecular weight / Extinction coefficient / pI - of protein</b>
	9.26 kDa / 12,490 M <sup>-1</sup> cm <sup>-1</sup> / 6.01
<b>5</b>	<b>Comments on sequence of expressed construct</b>
	N-terminal "WSHPQFEKGGG" eleven artificial residues due to construct design.
<b>6</b>	<b>Feeding buffer</b>

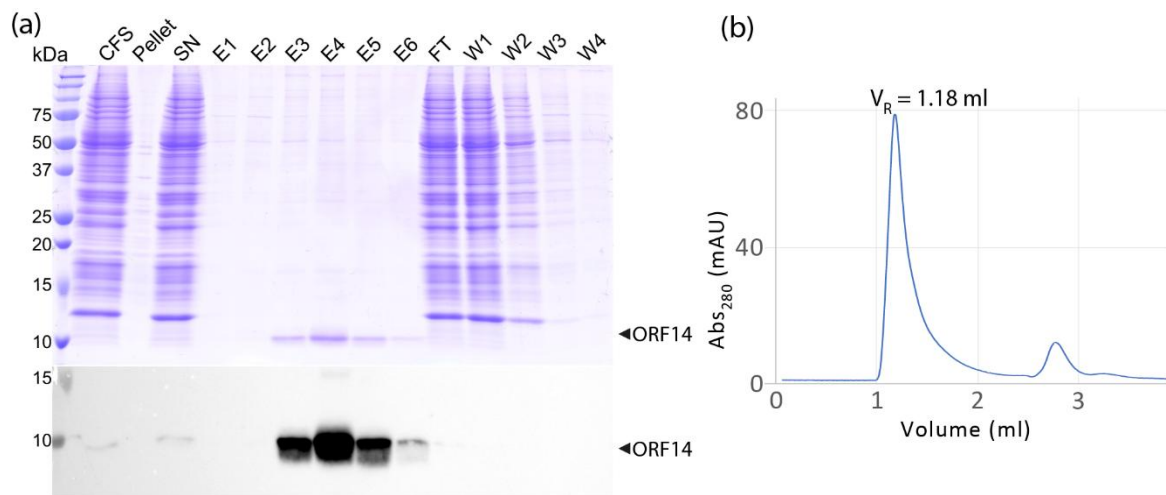
	30 mM HEPES-KOH (pH 7.6), 100 mM potassium acetate, 2.7 mM magnesium acetate, 16 mM creatine phosphate, 0.4 mM spermidine, 1.2 mM ATP, 0.25 mM GTP, 4 mM DTT and 6 mM (average concentration) amino acid mix and 0.05% (w/v) Brij-58.
<b>7</b>	<b>Translation mix</b>
	50% (v/v) mRNA, 50% (v/v) home-made WGE, 40 µg/mL creatine kinase, and 6 mM (average concentration) amino acid mix 0.05% (w/v) Brij-58.
<b>8</b>	<b>Protein synthesis temperature and time</b>
	22°C for 16 h without agitation (bilayer method).

Table 3: Protein Purification

<b>1</b>	<b>Buffer List</b>
A	100 mM Tris-HCl (pH 8.0), 150 mM NaCl, 1 mM EDTA, 0.1% (w/v) DDM (wash buffer).
B	100 mM Tris-HCl (pH 8.0), 150 mM NaCl, 1 mM EDTA, 2.5 mM desthiobiotin, and 0.1% (w/v) DDM (elution buffer).
<b>2</b>	<b>Purification steps (with corresponding buffer(s) and incubation times)</b>
A	Harvest total CFS.
B	Incubate with benzonase for 30 min on a wheel, at rt.
C	Centrifuge for 30 min at 20,000 g, 4°C.
D	Harvest the soluble fraction (SN).
E	Equilibrate the Strep-Tactin column (IBA Lifesciences) with 2 CV of <b>1A</b> (all steps performed on the bench by gravity).
F	Load SN onto the column.
G	Wash the column with 5 CV of <b>1A</b> .
H	Elute the protein of interest with <b>1B</b> .

Table 4: Final sample

<b>1</b>	<b>Yield</b>
	0.43 mg/mL WGE
<b>1b</b>	<b>A260/280 ratio</b>
	1.06
<b>2</b>	<b>Stability</b>
	protein has proved unstable during lipid insertion using cyclodextrin for detergent removal
<b>3</b>	<b>Comment on applicability</b>
	Solution NMR shows severely broadened resonances hinting to oligomerization or too big protein micelles. Lipid reconstitution is ongoing.



**(a) WG-CFPS in presence of detergent and Strep-tag purification of ORF14.** SDS-PAGE (upper panel) and WB (lower panel). **(b) SEC profile of ORF14.**