

## SI5: Nsp7

Table 1: General Information

<b>1</b>	<b>Protein Name (according to NCBI Reference Sequence NC_045512.2)</b>
	ORF1a and ORF1ab; nsp7
<b>2</b>	<b>Region/Name/Further Specification</b>
	Nsp7
<b>3</b>	<b>Sequence of fl protein (according to NCBI Reference Sequence NC_045512.2)</b>
	SKMSDVKCTSVVLLSVLQQLRVESSSKLWAQCVQLHNDILLAKDTTEAFEKMSVLLSVLLSMQ GAVDINKLCEEMLDNRATLQ
<b>4</b>	<b>Protein boundaries of expressed construct (according to NCBI Reference Sequence NC_045512.2)</b>
	aa 1-83 (fl nsp7)
<b>5</b>	<b>Ratio for construct design</b>
	fl protein
<b>6</b>	<b>Sequence homology (to SCoV)</b>
	Identity: 98.8%; similarity: 100%
<b>7</b>	<b>Published structures (SCoV2 or homologue variants)</b>
	SCoV: PDB 2KYS, 1YSY, 6NUS, 6NUR, 2AHM, SCoV2: PDB 7BV2, 7BV1, 6YYT, 7BTF, 6WQD, 6WTC, 6WIQ, 6M71, 6YHU, 6XEZ, 6M5I, 7CTT, 7C2K, 7BW4, 7BZF, 7JLT, 7AAP, 6XIP, 6XQB
<b>8</b>	<b>(Published) assignment (SCoV2 or homologue variants)</b>
	SCoV: PDB 1YSY, BMRB 6513, PDB 2KYS, BMRB 16981 SCoV2: BMRB 50337

Table 2: Protein Expression

<b>1</b>	<b>Expression vector</b>
	pET46
<b>2</b>	<b>Purification-/Solubility-Tag</b>
	N-terminal His <sub>6</sub> , enterokinase
<b>3</b>	<b>Cleavage Site</b>
	TEV
<b>4</b>	<b>Molecular weight / Extinction coefficient / pI - of cleaved protein</b>
	9.24 kDa / 5500 cm <sup>-1</sup> M <sup>-1</sup> / 5.2
<b>5</b>	<b>Comments on sequence of expressed construct</b>
	N-terminal "G" an artificial residue due to TEV-cleavage.

<b>6</b>	<b>Used expression strain</b>
	<i>E. coli</i> Rosetta2 pLysS
<b>7</b>	<b>Cultivation medium</b>
	M9 (uniformly <sup>15</sup> N-, <sup>13</sup> C-labelled)
<b>8</b>	<b>Induction system</b>
	IPTG inducible T7 promoter
<b>9</b>	<b>Induction of protein expression</b>
	0.5 mM IPTG at OD <sub>600</sub> 0.8
<b>10</b>	<b>Cultivation temperature and time</b>
	16°C for 14-16 h

Table 3: Protein Purification

<b>1</b>	<b>Buffer List</b>
A	10 mM HEPES (pH 7.4), 300 mM NaCl, 30 mM imidazole, 2 mM DTT.
B	10 mM HEPES (pH 7.4), 300 mM NaCl, 300 mM Imidazole, 2 mM DTT.
C	10 mM MOPS (pH 7.0), 150 mM NaCl, 2 mM DTT.
<b>2</b>	<b>Purification steps (with corresponding buffer(s) and incubation times)</b>
A	Cell lysis in buffer <b>1A</b> by microfluidizer operating at 20,000 psi. Lysates were cleared by centrifugation at 25,000 g for 30 min and then filtered through a 0.45 µm filter. Ni-NTA Agarose beads (Qiagen) were added to cleared lysates and incubated for 30 min. Beads were collected by centrifugation and then loaded onto a gravity column. Beads were washed twice with 10 column volumes of buffer <b>1A</b> . Protein was eluted with 5 column volumes of buffer <b>1B</b> .
B	Eluted protein was cleaved with 1% (w/w) TEV protease o.n. at rt while dialyzing the protein into 1 L buffer <b>1C</b> . Uncleaved protein was removed by inv. Ni-NTA binding.
C	Protein was concentrated using a 10 kDa MWCO (Amicon) concentrator and purified on aSD 200 Increase 10/300 (GE Life Sciences) size exclusion column, AKTApure (GE Life Sciences) using buffer <b>1C</b> .

Table 4: Final sample

<b>1</b>	<b>Yield</b>
	17 mg/L <sup>13</sup> C, <sup>15</sup> N-M9 medium
<b>1b</b>	<b>A260/280 ratio</b>
	0.5
<b>2</b>	<b>Stability</b>
	NMR sample stable at 4°C for a month, at 35°C for several days before degradation occurs.
<b>3</b>	<b>Comment on applicability</b>

	Suitable for NMR-based screening applications.
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